

Extreme and rapid bursts of functional adaptations shape bite force in amniotes

Article

Accepted Version

Sakamoto, M., Ruta, M. and Venditti, C. (2019) Extreme and rapid bursts of functional adaptations shape bite force in amniotes. *Proceedings of the Royal Society B: Biological Sciences*, 286 (1894). p. 20181932. ISSN 0962-8452 doi: <https://doi.org/10.1098/rspb.2018.1932> Available at <https://centaur.reading.ac.uk/81500/>

It is advisable to refer to the publisher's version if you intend to cite from the work. See [Guidance on citing](#).

To link to this article DOI: <http://dx.doi.org/10.1098/rspb.2018.1932>

Publisher: The Royal Society

All outputs in CentAUR are protected by Intellectual Property Rights law, including copyright law. Copyright and IPR is retained by the creators or other copyright holders. Terms and conditions for use of this material are defined in the [End User Agreement](#).

www.reading.ac.uk/centaur

CentAUR

Central Archive at the University of Reading

Reading's research outputs online

RUNNING HEAD: Bite force evolution in amniotes

Extreme and rapid bursts of functional adaptations shape bite force in amniotes

Manabu Sakamoto^{1*}, Marcello Ruta², Chris Venditti^{1*}

¹School of Biological Sciences, University of Reading, Reading, Berkshire, RG6 6BX, UK

²School of Life Sciences, University of Lincoln, Lincoln, Lincolnshire, LN6 7DL, UK

***Corresponding authors:**

Manabu Sakamoto

School of Biological Sciences, University of Reading, Reading, UK, RG6 6BX

m.sakamoto@reading.ac.uk

Chris Venditti

School of Biological Sciences, University of Reading, Reading, UK, RG6 6BX

c.d.venditti@reading.ac.uk

ABSTRACT

Adaptation is the fundamental driver of functional and biomechanical evolution. Accordingly, the states of biomechanical traits (absolute or relative trait values) have long been used as proxies of adaptations in response to direct selection. However, ignoring evolutionary history, in particular ancestry, passage of time and the rate of evolution, can be misleading. Here, we apply a recently developed phylogenetic statistical approach using significant rate shifts to detect instances of exceptional rates of adaptive changes in bite force, in a large group of terrestrial vertebrates, the amniotes. Our results show that bite force in amniotes evolved through multiple bursts of exceptional rates of adaptive changes, whereby whole groups – including Darwin’s finches, maniraptoran dinosaurs (group of non-avian dinosaurs including birds), anthropoids and hominins (the group of species including modern humans) – experienced significant rate increases compared to the background rate. However, in most parts of the amniote tree of life we find no exceptional rate increases, indicating that coevolution with body size was primarily responsible for the patterns observed in bite force. Our approach represents a template for future studies in functional morphology and biomechanics, where exceptional rates of adaptative changes can be quantified and potentially linked to specific ecological factors underpinning major evolutionary radiations.

KEYWORDS

Bite force, evolution, phylogenetic comparative methods, adaptations, amniotes, rate-shifts

BACKGROUND

Adaptation is the fundamental driver of functional and biomechanical evolution. Measures of biomechanical performance – e.g. bite force – characterize and quantify specific functional performances to fulfil ecological demand – e.g., diet [1, 2]. Functions are therefore typically assumed to be under direct selection – i.e. a change in biomechanical performance indicates selection for changes in function. For instance, taxa with higher bite force are often interpreted as having diets requiring powerful bites, driving selection on associated morphological features [3-5]. However, observable trait states can be the result of multiple confounding factors independent of direct selection for a specific function, including scaling effects [6] and evolutionary history (ancestry and the passage of time). That is, species can often achieve relatively high bite forces owing to the accumulation of evolutionary changes that is within expectation given the passage of time, without invoking exceptionally strong selection pressures towards increased bite forces. Assuming that the states of biomechanical traits can be used as proxies for strong selection/functional demand while ignoring such confounding factors (in particular evolutionary history) can be and often are misleading [7, 8]. Thus, it is important to identify cases in which trait values exceed the amount of evolutionary change expected given confounding factors – i.e., objectively detecting instances of *exceptional rates of adaptive changes* [9].

It has long been interpreted that the intensity of natural selection acting on a phenotype is linked to rates of evolution [9, 10], and significant shifts in rates can be interpreted as instances of adaptive responses to strong selective pressures – e.g., positive phenotypic selection [9] akin to positive genetic selection [11, 12]. Positive selection is invoked as an explanation for trait evolution along a branch on a phylogenetic tree if the amount of evolutionary change exceeds the amount of change expected from the passage

of time given the background rate of evolution. In the context of biomechanical evolution, evidence for exceptional rates of adaptive changes in biomechanical traits can be detected as significant rate shifts using this phylogenetic comparative framework. However, in spite of the ever-increasing numbers of comparative biomechanical studies employing a phylogenetic framework [13-27], studies attempting to detect episodes of exceptional rates of adaptive changes in biomechanical traits that can be interpreted as evolutionary responses to strong levels of natural selection using a statistically rigorous comparative framework are still comparatively rare [but see, e.g., 23].

Here, we test the hypothesis that instances of exceptional rates of adaptive changes have shaped the observed diversity in biomechanical traits focusing on bite force evolution. Our hypothesis specifically relates to detecting exceptional rates of adaptive changes and not in detecting rate heterogeneity – exceptional rates of adaptation is defined as instances of exceptionally large rate increases (at least twice the background rate, see Methods and [9] for details). Bite force relates to species' niche and feeding ecology, is correlated with several ecological and behavioural traits [28-31], and is widely available from the literature across several fields of study (e.g., biomechanics, ecology, palaeobiology) for a broad sample of the tree of life, making it an ideal biomechanical trait to test our hypothesis using a phylogenetic framework. To this end, we assembled the largest dataset of bite forces collected to date for amniotes, both extinct (including non-avian dinosaurs, sabre-toothed cats, and hominins) and extant.

METHODS

Data. Bite force and body mass data were collected primarily through the literature, augmented with novel estimates (ESM), spanning 434 extant and extinct amniote species

(Table S1). We used the Time Tree of Life (TTOL) [32] as the backbone phylogeny, with fossil tips/clades inserted at the appropriate phylogenetic and temporal positions (ESM) using fossil dates from the Paleobiology Database (accessed 9 Feb 2017).

Variable-Rates Phylogenetic Regression Models. We fitted a variable-rates (VR) regression model [9] using BayesTraits [33] on \log_{10} bite force against \log_{10} body mass (*Single-slope VR model*). The VR regression model as implemented in BayesTraits works to modify the branch lengths to “detect heterogeneity in rates of phylogenetically structured residual errors” [9]. That is, once the appropriate level of variance in the response variable – e.g., bite force – is explained by some predictor variable(s) – e.g., body mass – outlying deviations from the regression line will be explained as rate shifts (Fig. S1). Under Brownian motion, bite force – after accounting for body mass and other confounding variables – evolves at a rate proportional to time (and an estimated background variance) across the phylogeny, and for any evolutionary change along a given branch that is greater/less than the expected amount of change for the duration of time to occur (given body mass), that branch must be stretched/compressed in length in proportion to the observed amount of phenotypic change – corresponding to a rate increase/decrease. The magnitude of branch stretching/compressing is the rate scalar (r).

As previous research indicated that scaling of bite force is group-specific [34-36] we tested an additional model in which separate slopes were estimated for five different groups (*5-Group VR model*). Each taxon was assigned to one of five groups: mammals excluding bats (hereafter “Mammals”), bats, finches (Fringillidae, Estrildidae, and Darwin’s finches); non-finch dinosaurs (including other birds, hereafter “Dinosaurs”); and non-dinosaurian diapsid reptiles (hereafter “Reptiles”) (Fig. 1; Table S1). We chose these five

groups because they act as good descriptors of the distribution of data (Fig. S3) as well as conforming to widely recognized taxonomic groups (see ESM for details). Birds and dinosaurs were grouped together, as fossil evidence points to a blurred distinction in their physiology and biology [37-40], while dinosaurs are very different from other reptiles [41]. Intercept differences were not modelled – group-wise offsets will trade off with rate scalars on branches leading to the last common ancestors of the respective groups (Fig. S1). In order to account for potential differences owing to bite force acquisition type and biting positions (ESM), two additional models were fitted with the confounding variables Bite Type and Bite Point individually added to the regression model as covariates (ESM).

We tested for instances of exceptional bursts of evolutionary change in bite force based on rate shifts along branches on the phylogeny [9], based on the premise that phenotypic changes owing to adaptations (potentially as a response to strong selective pressure) would be proportional to r . Thus, we define exceptional change following the criteria of [9]: 1) certainty of rate shifts, the branch in question must be scaled in >95% of the posterior sample of scaled trees; and 2) magnitude of rate shifts, the r in question must be greater than two. Rate heterogeneity that do not fulfil these two criteria were not considered as instances of exceptional rates of adaptive changes. There are two types of rate shifts: branch-wise rate shifts (branch shifts), are cases in which significant increases in rates with respect to background rate are detected along individual branches; and clade-wise rate shifts (clade shifts), occur across all branches within a clade and represent cases in which rapid divergences in trait values have occurred (Fig. S1). We determined whether rate-shifts constituted exceptional rates of adaptive changes if they satisfied the criteria set out above in all of three independent replicate Markov Chain Monte Carlo (MCMC) chains.

In order to determine if rate-heterogeneity was statistically significant, we fitted an *equal-rate (ER) model* (or Brownian motion) as a simpler alternative to each of our VR models. Model selection was performed using the Bayes Factor (BF; see ESM for details): BF is defined as twice the difference in log marginal likelihood (m) between the complex model (model_1) and the simple model (model_0) – i.e., $\text{BF} = 2 \times (m_1 - m_0)$. For instance, we computed BF using m from our 5-Group VR model and the simple alternative 5-Group ER model, and selected the VR model over the ER model when BF value was greater than 2 [42].

We ran our MCMC chains for 10^9 iterations, with a burn-in period of 10^8 iterations, sampling every 10^5 iterations, resulting in a posterior sample of 900 modified VR trees and model estimates, for each regression model. We used stepping stone sampling (over 1000 stones at 10^5 iterations each) to compute marginal likelihoods from which BF were calculated. Post-processing of the BayesTraits outputs were conducted using an online post-processor (www.evolution.reading.ac.uk/VarRatesWebPP), as well as in R [43].

RESULTS

Variable-Rates Regression Model. We found strong support for the VR model compared to the ER model ($\text{BF}_{\text{VR-ER}} = 474$) for the single-slope regression model. Bite force scales nearly isometrically with body mass, with a slope of 0.674 ($p\text{MCMC}_0 < 0.001$, $R^2_{\text{mean}} = 0.79$; Table S2), which is not significantly different from a theoretical isometric slope of 0.67 ($p\text{MCMC}_{0.67} = 0.4$) [6].

There is statistical support for favouring the 5-Group model over the single-slope model (Fig. 1)); significant differences exist among the slopes of different groups (Table S2; Table S3). Finches and bats are not different from each other, but are distinct from

mammals, reptiles and dinosaurs (Fig. 1; Table S3); in turn, these three groups are not different from each other (Fig. 1; Table S3). Finches and bats have slopes that deviate from 0.67 (Table S4), while the other three groups have slopes that are not significantly different from 0.67 (Table S4). Critically, despite allowing for the variation in slopes among taxonomic groups, our 5-Group VR model ($R^2_{\text{mean}} = 0.809$; Table S2) still outperforms a 5-Group ER model ($\text{BF}_{\text{VR-ER}} = 429$; Table S2).

Model selection showed that bite type is not significant ($p\text{MCMC} > 0.05$ in both Single-Slope+BiteType and 5-Group+BiteType models; Table S2) while bite point is ($p\text{MCMC} < 0.05$ in both Single-Slope+BitePoint and 5-Group+BitePoint models; Table S2). Given a similar body size, bite force is comparable in magnitude between *in vivo* measurements and indirect estimates, but it differs in magnitude between posterior and anterior bites (posterior positions have higher forces, as expected). There is no slope difference between bite type categories or between bite point categories (Figs S6, S7). We used the 5-Group+BitePoint model as our final model for detecting exceptional rates of adaptive changes in bite force – this enables us to compare evolutionary rates after accounting for effects owing to body size and bite point.

Rate Shifts and Exceptional Rates of Adaptive Changes. We found substantial amount of rate heterogeneity (elevated rates in >50% of the posterior sample) in the amniote tree of life, along 439 branches out of 866 branches in the phylogeny (51% of branches; Fig. S4). Instances of exceptional rates of adaptive changes are found in a far fewer number of branches: in 182 branches (21% of branches) (Table S5; Figs 1, 2a, S5; Movie S1). Our results show that bite force evolved through multiple bursts of exceptional rates of adaptive changes, whereby whole groups experienced rate increases of bite force evolution

compared to the background rate across the entire amniote tree. We find such clade shifts in Darwin's finches (median $r > 55$), the hominin lineage excluding *Australopithecus anamensis* ($r > 35$), Anthroidea ($r > 6$), and maniraptoran theropod dinosaurs (the clade including birds and their closest relatives, here *Erlikosaurus* and *Dromaeosaurus*; median $r > 3$). Thus, most of the scaled branches are because of a smaller number of node scalars that modify all descendant branches.

Two aspects of the distribution of rate shifts are noteworthy. First, clade-wise rate shifts show a nested pattern (Fig. 1). For example, Darwin's finches exhibit an additional level of rate increase above that of the maniraptoran rate increase (Figs 1, 2a, 2b, S5; Movies S1-S2). The same pattern characterizes hominins (excluding *A. anamensis*) within anthropoid primates (Figs 1, 2a, 2c, S5; Movies S1-S2). In particular, *Homo* species exhibit reductions in bite forces (Fig. 2c), which is in marked contrast to the apparent increase in hominin body size through time (Figs S8) [44]. Thus, humans drastically reduced bite force through time at a rate faster than their anthropoid ancestors and relatives.

The second key aspect of the rate shift distribution is that branch-wise rate increases occur in conjunction with clade-wide shifts. We identified an exceptional increase in the rate of bite force evolution along the branch leading to Passeroidea (the clade defined by the last common ancestor of our finches) (median $r > 30$), followed by a reversal to the ancestral maniraptoran rate. We also recovered a scattering of branch-specific shifts on terminal branches (*Proteles cristatus*, *Panthera onca*, *Sus scrofa*, *Stegosaurus*, and *Plateosaurus*), marking sudden changes in the biomechanical performance of some species from their close relatives (Figs 1, 2a, S5; Movie S1).

DISCUSSION

Exceptional Rates of Adaptive Changes in Bite Force. Working under the premise that the rate of phenotypic trait evolution is proportional to the strength of selection [9, 10], we provide the first evidence for exceptional rates of adaptive changes shaping the diversity of bite force in both extinct and extant amniotes, using a statistically robust evolutionary framework [9]. We find such instances of exceptional rates of adaptive changes in four clades of amniotes and a handful of independent branches. Conversely, in most lineages of amniotes studied here (79% of branches in the phylogeny), bite force does not undergo exceptional rates of adaptive changes (even in lineages with elevated rates; ESM; Fig. S4), indicating that co-evolution with body size is, for the most part, the main factor responsible for bite force variation. A large predator can generate enough bite force to kill its prey just by being large. As an example, *Tyrannosaurus rex* was most likely capable of “pulverizing” bones [45] simply owing to its colossal size (~5-10 tonnes [46-48]). We did not detect instances of exceptional rates of adaptive changes in bite force in this taxon, and therefore there is no evidence of strong selection for a feeding ecology that requires disproportionately high bite force – e.g. “extreme osteophagy (bone consumption)” [45]. Similarly, we do not detect signatures of exceptional rates of adaptive changes in classically recognised power-biters such as osteophagous hyenids [49], short-faced hyper-carnivorous felids [50, 51], and small-brained carnivorous marsupials [3, 5, 52] indicating that bite force in these clades were not subjected to strong selection as is often presumed.

Interestingly, contrary to our prior expectations, we do not detect exceptional rates of adaptive changes in sabre-toothed cats (Machairodontinae). Although rates are on average higher in Felidae as a whole (including both sabre- and conical-toothed cats along with the basal cats *Proailurus* and *Hyperailurictis*) compared to the background rate in the majority of the posterior sample (i.e., >50%; ESM; Fig. S4), they do not fulfil the criteria for

exceptional rates of adaptive changes. Further, there is no difference between rates in machairodontine lineages and other felid lineages. The time elapsed in the lineage leading to sabre-toothed cats since their divergence with conical-toothed cats sufficiently explains the reduction in bite force in sabre-toothed cats.

It is possible that for the majority of taxa in our bite force sample, individuals preferentially seek out and consume food items that can be processed within the naturally generated range of bite forces, and rarely actively seek food items that require maximum biting capacity. This equates to a behavioural adaptation, in which species evolve bite force through correlated evolution with body mass, and preferentially consume food items that fit within their natural range of bite force. If this is true, then selection for improved biting performance with respect to higher bite force may not frequently occur. Additionally, functional-morphological adaptations – e.g. tooth morphology – may facilitate higher biting performance – e.g., piercing, crushing or shearing – without necessitating a more powerful bite.

Similarly, drastic reduction in bite force – as a trade-off between force and velocity if jaw closing velocity was under strong selection – would constitute an exceptional rate of adaptation significantly below the expected range of bite force given the universal scaling relationship. However, it is potentially more likely for reductions in bite force to occur over exceptional gains in bite force since muscles are expensive organs to maintain and more so to enlarge. Indeed, we observe more instances of drastic reductions in bite force associated with exceptional rates of adaptive changes than we do exceptional increases in bite force (Figs 2, S5).

Despite the overall uniform evolution of bite force relative to body mass, we find evidence for exceptional rates of adaptive changes playing a major role in the evolution of

bite force (just not as a tree-wide pattern across all major clades). Since the majority of these exceptional rates of adaptive changes occur as clade-wide rate shifts, it is possible that they are linked to some biological, ecological or environmental features unique to those clades and shared amongst constituent members. For instance: the acquisition of a “key” innovation, which allows such clades to rapidly expand and exploit functional niches; a shift into a new environment, habitat or lifestyle that is associated with new opportunities and resources; or an extrinsic environmental event (such as mass extinction events) that results in an abundance of ecological niches available for exploitation. Determining such factors ultimately responsible for exceptional rates of adaptive changes is theoretically possible. Namely, the VR regression framework allows for the inclusion of extrinsic factors such as dietary preference, feeding strategy, sexual display/conflict, etc., as additional covariates. A covariate can be identified as the extrinsic driver of bite force evolution if its inclusion can explain much of the variation in bite force, thereby reducing or eliminating rate shifts. At present, ecological data associated with biting performance are only available for a handful of species, but we hope that future work will considerably augment information on ecological covariates.

Macroevolutionary Patterns of Exceptional Rates of Adaptive Changes. Overall, our results highlight a combination of clade-wise and branch-wise rate shifts occurring across the amniote phylogeny. Clade-wise rate shifts are characterized by an elevated rate that is homogenous across all branches within a given clade and are associated with an increase in trait variation in the constituent taxa given the variance in the other taxa in the data [9]. Lineages in such clades continually evolve traits at a faster rate through time compared to other parts of the tree (Fig. 1). This contrasts with a classic description of an adaptive

radiation [53], which is characterized by a rapid initial burst of trait evolution followed by a rate slowdown associated with niche saturation [54]; clade-wise rate shifts see continual changes in functional niche occupation. One implication of such patterns in bite force evolution is that evolutionary lineages (sequence of branches leading to terminal taxa) do not stay in the same regions of function-space (pertaining to biting functional variation), but rather, continue to expand out to unoccupied/unexplored regions of function-space. Evolutionary lineages will be moving through various functional niches as their bite force values change through time. An alternative interpretation is that function-space itself changes through time. i.e. functional/ecological niches are dynamic rather than fixed entities, a constantly moving target [55]. Yet another interpretation is that function-space saturates but convergences occur frequently and rapidly – that is, lineages move in and out of occupied/explored regions of function-space. In this context, our results would support the notion that functional adaptations are relatively labile over evolutionary history and remain responsive to changing environmental conditions and ecological demands.

Branch-wise rate shifts (rate shifts associated with single branches only) occurring on branches subtending whole clades (Fig. 1), such as that observed at the base of Passeroidea in our dataset, can be interpreted as a mean-shift in bite force after accounting for body mass [9]. In our case, this means that there was a rapid shift in the mean bite force value of Passeroidea from the ancestral maniraptoran mean. The total sum of evolutionary changes accumulating along the branch leading to Passeroidea exceeds that expected from the temporal duration of that branch. This is irrespective of any un-sampled taxa along the lineage – e.g., other perching birds (Passeriformes, e.g. corvids, shrikes) for which bite force data are not available in the literature as far as we are aware.

Similarly, the two large-bodied herbivorous taxa, *Stegosaurus* and *Plateosaurus*, have evolved bite force at excessively high rates (~11 and ~35 times background rate, respectively – *Stegosaurus* and *Plateosaurus* have extremely small heads, and thus low bite forces, for their body sizes), but these could potentially represent evolutionary patterns within thyreophoran and sauropodomorph dinosaurs respectively, and not specifically associated with these two species. Using a different measure of size such as head length or width may likely change these results – though, body mass has major benefits over head size for its ecological implications. Nonetheless, major changes in bite force relative to body mass did occur along these lineages so the interpretation remains the same: the amount of change in trait values given the duration of time elapsed is exceptionally high compared to the background rate.

Evolution of Bite Force in Maniraptoran Dinosaurs. Maniraptoran theropods are perhaps the most diverse amongst dinosaurs in terms of functional and morphological specializations associated with feeding. Forms like the parrot-like oviraptorosaurs, large herbivorous therizinosaurs, hyper-carnivorous dromaeosaurs with recurved teeth (e.g. *Velociraptor*), and toothed and toothless avialans are just some typical examples of maniraptoran morpho-functional diversity. High evolutionary rates in maniraptoran bite force indicate that their morphological and presumed ecological diversity are linked with selection on biting performance. Maniraptoran fossils are predominantly known from Cretaceous rocks but are inferred to have originated by the Middle Jurassic (~168 Myr ago; Fig. 1), with derived members including the avialan *Archaeopteryx* appearing relatively quickly, by the Late Jurassic (~150 Myr ago; Fig. 1). This implies that Maniraptora underwent a rapid diversification (both in species diversity but also in bite force variance) early in their

evolutionary history, but that they retained high evolutionary rates in bite force throughout the clade's history.

The observation that bite force underwent exceptional rates of adaptive changes in maniraptoran theropods but not uniquely in birds – rates in birds are not distinguishable from those in other non-avian maniraptorans – is consistent with recent findings that the evolution of birds and their immediate close relatives – i.e. paravians – are similar to one another [37], and that many of the features traditionally associated with birds were present in paravians and more broadly in maniraptorans. Here we have demonstrated that this is also the case with bite force evolution (given the available data); the rate of bite force evolution did not change from non-avian maniraptorans to birds. On the other hand, this means that heritable rates of bite force evolution in maniraptoran ancestors possibly contributed to some extent on the subsequent ecological success of birds – the ability to rapidly change bite force in response to changing environmental and ecological pressures would surely have been beneficial for early Cenozoic birds in the post-extinction world.

Our identification of an extreme clade-wide rate shift in the Darwin's finches, which is among the highest in the tree (>55 times the background rate; Figs 1, 2, S5; Movies S1-S2; Table S5), is noteworthy for both historical and biological reasons. Darwin's finches are the classic textbook case of 'adaptive radiation', with eco-morphological diversification occurring in a short time interval after the initial colonization of the Galapagos Islands by finches [56-58]. Their diversification in feeding ecology is particularly relevant to the rapid evolution of bite force, as Darwin's finches are well documented to have strong dietary preferences on food types of varying toughness [59] or differences in food manipulations [60]. Within the context of the evolutionary history of amniotes as sampled here (~350 Myr), the radiation of Darwin's finches is comparatively recent with some divergences

occurring in a geologically instantaneous manner (Fig. 1). Compared to their recent divergence times, bite force variance in Darwin's finches is exceptionally high spanning almost two orders of magnitude (Fig. 2) accounting for their extraordinarily high evolutionary rates.

Evolution of Bite Force in Humans. The exceptionally high rates of bite force evolution in the hominins excluding *A. anamensis* (Figs 1, 2) – more than 35 times the background rate and ~6 times those for the branches within other anthropoid primates – highlights an important, recent and rapid evolution in our own lineage. In particular, the decrease in bite force in *Homo* species (Fig 2) is contrary to the increase in hominin body size through time (Fig S8) – such a discrepancy between bite force and body mass is indicative of strong directional selection, and coincides with previously documented evolutionary shifts in relative molar sizes, attributed to the reduction in feeding time associated with the introduction of food processing such as cooking [9, 25]. Strikingly, [61] found a rate shift in symphyseal angle in the hominin lineage excluding *A. anamensis*, coinciding with the phylogenetic location where we find a rate shift in bite force (Fig. 1).

Additionally, the reduction in bite force in the hominin lineage may have occurred as a consequence of an evolutionary trade-off with increasing brain size in this group [62-64] (see ESM; Figs S9-S10). As brain size increases relative to skull size, the temporal fossa (defined as the opening between the braincase and the zygomatic arch) is reduced in dimension, thereby decreasing the amount of space available to house the temporal muscles [63, 64], which are critical for achieving hard biting in most animals. This reduction in temporal muscle can be seen in the changing predominance of the sagittal crest (ridge of bone running along the midline of the skull) where the temporal muscles attach through

hominin evolution (Fig. S10). Strikingly, molecular evidence supports the hypothesis that a drastic reduction in the temporal muscle occurred along the lineage to *H. sapiens* after its divergence from chimpanzees owing to a frameshifting mutation, causing inactivation of the predominant myosin heavy chain expression in masticatory muscles [63]. The mutation has been inferred to have coincided approximately with the enlargement of the brain, presumably concurrent with the origin of the genus *Homo* [63] – though the timing has been contested [65, 66]. Furthermore, *H. sapiens* relies less on the temporal muscles and more on the masseter muscles to generate bite force [67]. Thus, such a trade-off between temporal muscle size and brain size is a reasonable explanation for the evolutionary reduction in bite force in the hominin lineage through time, with the advent of cooking further accelerating the loss of reliance on high bite force for food processing.

Indeed, an auxiliary phylogenetic regression modelling [68] of bite force on brain size (endocranial volume) [69-71] accounting for body mass on hominids (*Pongo*, *Gorilla*, *Pan*, and hominins), shows that bite force scales negatively with brain size (ESM; Fig. S9). The reduction in bite force is statistically associated with an increase in brain size.

Alternatively, the reduction in bite force may owe to neoteny, whereby the jaw muscles arrest in development along the hominin lineage. However, neoteny in humans has also been interpreted as being associated with brain enlargement so the precise cause of the reduction in jaw adductor musculature and thus bite force is up for debate.

CONCLUSIONS

Taken together, our results reveal that the evolution of bite force in amniotes occurred as bursts of accelerated changes across multiple clades, and as the product of repeated and nested pulses of progressively higher rates of change, representing instances of exceptional

400 functional adaptations. Using a phylogenetic evolutionary framework, on a dataset
401 representing the largest taxonomic sample to date, enables us to statistically detect
402 instances of adaptations in biomechanical metrics, not only along specific branches, but also
403 through time, paving the way to better understand how specific ecological niches (feeding
404 ecologies) are occupied. In order to determine whether species' bite force underwent
405 instances of exceptional rates of adaptive changes, it is necessary to demonstrate an
406 exceptionally high rate of evolution associated with that species in a phylogenetic context
407 after accounting for size and the expected evolutionary change associated with divergence
408 time.

409 **DATA ACCESSIBILITY**

410 Data are available on the Dryad Digital Repository (provisional DOI:

411 doi:10.5061/dryad.q12c06f). Temporary review link is as follows:

412 <https://datadryad.org/review?doi=doi:10.5061/dryad.q12c06f>

413

414 **ETHICS STATEMENT**

415 Not applicable

416

417 **COMPETING INTERESTS**

418 We have no competing interests.

419

420 **FUNDING**

421 MS and CV were supported by the Leverhulme Trust (RPG-2013-185 and RPG-2017-071) and

422 MS and MR were supported by the BBSRC (BB/H007954/1).

423

424 **AUTHOR CONTRIBUTIONS**

425 MS, MR and CV conceived of the study, designed the study, coordinated the study and draft

426 the manuscript. MS collected and analysed the data. All authors gave final approval for

427 publication.

428

429 **ACKNOWLEDGMENTS**

430 We thank Joanna Baker, Ciara O'Donovan, Andrew Meade, Jorge Avaria Llatureo, and

431 Henry Ferguson-Gow for invaluable discussion, which helped improve the research and

432 manuscript. MS thanks the curators and collection managers at the various institutions

433 visited as part of the data collection process, in particular Rhian Rowson (Bristol City
434 Museum and Arts Gallery), Paolo Viscardi (National Museum of Ireland – Natural History;
435 formerly Horniman Museum), Mark Carnall (Oxford University Museum; formerly Grant
436 Museum, UCL), Lars Werdelin (Swedish Museum of Natural History), Andrew Kitchener
437 (National Museums Scotland – Natural Sciences), Judy Galkin (American Museum of Natural
438 History), Roberto Portela Miguez (Natural History Museum, London), and Milly Farrell
439 (Oxford Brookes University; formerly Hunterian Museum, Royal College of Surgeons). We
440 also thank Larry Witmer (Ohio University) for permission to use his gallery of various
441 theropod skull images. Finally, we thank the reviewers for their constructive comments on
442 this manuscript. Silhouettes in Figs 1 and 2 are attributed to Andrew Farke (*Stegosaurus*, CC-
443 BY 3.0), Michael Keesey (*Homo sapiens*, CC-BY 3.0; *Gorilla*, CC0 1.0), Steven Traver
444 (*Sphenodon punctatus*, CC0 1.0), Sarah Werning (*Panthera tigris*, CC-BY 3.0) and Yan Wong
445 (Chiropteran, CC0 1.0). Anthropoid is uncredited (CC0 1.0). Silhouettes of finches were
446 drawn using photographs by Vince Smith (*Geospiza fuliginosa*, CC-BY 2.0) and Brian
447 Gratwicke (*Platyspiza crassirostris*, CC-BY 2.0). Silhouette of *Deinonychus antirrhopus* is
448 author's own work (MS, CC-BY 3.0).

REFERENCES

- [1] Grant, P.R. 1981 The feeding of darwin's finches on *Tribulus cistoides* (L.) seeds. *Animal Behaviour* **29**, 785-793. (doi:[http://dx.doi.org/10.1016/S0003-3472\(81\)80012-7](http://dx.doi.org/10.1016/S0003-3472(81)80012-7)).
- [2] Schluter, D., Price, T.D. & Grant, P.R. 1985 Ecological Character Displacement in Darwin's Finches. *Science* **227**, 1056-1059.
- [3] Wroe, S., McHenry, C. & Thomason, J. 2005 Bite club: comparative bite force in big biting mammals and the prediction of predatory behaviour in fossil taxa. *Proceedings of the Royal Society B-Biological Sciences* **272**, 619-625.
- [4] Christiansen, P. & Wroe, S. 2007 Bite forces and evolutionary adaptations to feeding ecology in carnivores. *Ecology* **88**, 347-358.
- [5] Wilson, G.P., Ekdale, E.G., Hoganson, J.W., Caledo, J.J. & Vander Linden, A. 2016 A large carnivorous mammal from the Late Cretaceous and the North American origin of marsupials. *Nat Commun* **7**, 13734. (doi:[10.1038/ncomms13734](https://doi.org/10.1038/ncomms13734) <http://www.nature.com/articles/ncomms13734#supplementary-information>).
- [6] Schmidt-Nielsen, K. 1984 *Scaling: Why is animal size so important?* Cambridge, Cambridge University Press.
- [7] Felsenstein, J. 1985 Phylogenies and the Comparative Method. *American Naturalist* **125**, 1-15. (doi:[Doi 10.1086/284325](https://doi.org/10.1086/284325)).
- [8] Harvey, P.H. & Pagel, M.D. 1991 *The comparative method in evolutionary biology*, Oxford University Press.
- [9] Baker, J., Meade, A., Pagel, M. & Venditti, C. 2016 Positive phenotypic selection inferred from phylogenies. *Biological Journal of the Linnean Society* **118**, 95-115. (doi:[10.1111/bij.12649](https://doi.org/10.1111/bij.12649)).

- 472 [10] Simpson, G.G. 1944 *Tempo and mode in evolution*. New York, Columbia University
473 Press; 237 p.
- 474 [11] Yang, Z. 2002 Inference of selection from multiple species alignments. *Current Opinion*
475 *in Genetics & Development* **12**, 688-694. (doi:http://dx.doi.org/10.1016/S0959-
476 437X(02)00348-9).
- 477 [12] Yang, Z. 2006 *Computational Molecular Evolution*. Oxford, Oxford University Press; 376
478 p.
- 479 [13] Ghalambor, C.K., Walker, J.A. & Reznick, D.N. 2003 Multi-trait Selection, Adaptation,
480 and Constraints on the Evolution of Burst Swimming Performance1. *Integrative and*
481 *Comparative Biology* **43**, 431-438. (doi:10.1093/icb/43.3.431).
- 482 [14] Walker, J., xa, A, Associate Editor: Troy, D. & Editor: Michael, C.W. 2007 A General
483 Model of Functional Constraints on Phenotypic Evolution. *The American Naturalist* **170**, 681-
484 689. (doi:10.1086/521957).
- 485 [15] Ackerly, D. 2009 Conservatism and diversification of plant functional traits: Evolutionary
486 rates versus phylogenetic signal. *Proceedings of the National Academy of Sciences* **106**,
487 19699-19706. (doi:10.1073/pnas.0901635106).
- 488 [16] Collar, D.C., O'Meara, B.C., Wainwright, P.C. & Near, T.J. 2009 Piscivory limits
489 diversification of feeding morphology in centrarchid fishes. *Evolution* **63**, 1557-1573.
490 (doi:10.1111/j.1558-5646.2009.00626.x).
- 491 [17] Martin, C.H. & Wainwright, P.C. 2011 Trophic novelty is linked to exceptional rates of
492 morphological diversification in two adaptive radiations of cyprinodon pupfish. *Evolution* **65**,
493 2197-2212. (doi:10.1111/j.1558-5646.2011.01294.x).

494 [18] Holzman, R., Collar, D.C., Price, S.A., Hulsey, C.D., Thomson, R.C. & Wainwright, P.C.
 495 2012 Biomechanical trade-offs bias rates of evolution in the feeding apparatus of fishes.
 496 *Proceedings of the Royal Society B: Biological Sciences* **279**, 1287-1292.

497 [19] Wainwright, P.C., Smith, W.L., Price, S.A., Tang, K.L., Sparks, J.S., Ferry, L.A., Kuhn, K.L.,
 498 Eytan, R.I. & Near, T.J. 2012 The Evolution of Pharyngognath: A Phylogenetic and
 499 Functional Appraisal of the Pharyngeal Jaw Key Innovation in Labroid Fishes and Beyond.
 500 *Systematic Biology* **61**, 1001-1027. (doi:10.1093/sysbio/sys060).

501 [20] Anderson, P.S.L., Claverie, T. & Patek, S.N. 2014 Levers and linkages: Mechanical trade-
 502 offs in a power-amplified system. *Evolution* **68**, 1919-1933. (doi:10.1111/evo.12407).

503 [21] Collar, D.C., Wainwright, P.C., Alfaro, M.E., Revell, L.J. & Mehta, R.S. 2014 Biting
 504 disrupts integration to spur skull evolution in eels. **5**, 5505. (doi:10.1038/ncomms6505
 505 <https://www.nature.com/articles/ncomms6505#supplementary-information>).

506 [22] Anderson, P.S.L. & Patek, S.N. 2015 Mechanical sensitivity reveals evolutionary
 507 dynamics of mechanical systems. *Proceedings of the Royal Society B: Biological Sciences* **282**.

508 [23] Muñoz, M.M., Anderson, P.S.L. & Patek, S.N. 2017 Mechanical sensitivity and the
 509 dynamics of evolutionary rate shifts in biomechanical systems. *Proceedings of the Royal*
 510 *Society B: Biological Sciences* **284**.

511 [24] Stayton, C.T. 2011 Biomechanics on the half shell: functional performance influences
 512 patterns of morphological variation in the emydid turtle carapace. *Zoology* **114**, 213-223.
 513 (doi:<https://doi.org/10.1016/j.zool.2011.03.002>).

514 [25] Organ, C., Nunn, C.L., Machanda, Z. & Wrangham, R.W. 2011 Phylogenetic rate shifts in
 515 feeding time during the evolution of Homo. *Proceedings of the National Academy of*
 516 *Sciences* **108**, 14555-14559. (doi:10.1073/pnas.1107806108).

517 [26] Piras, P., Sansalone, G., Teresi, L., Kotsakis, T., Colangelo, P. & Loy, A. 2012 Testing
518 convergent and parallel adaptations in talpids humeral mechanical performance by means
519 of geometric morphometrics and finite element analysis. *Journal of Morphology* **273**, 696-
520 711. (doi:10.1002/jmor.20015).

521 [27] Hagey, T.J., Uyeda, J.C., Crandell, K.E., Cheney, J.A., Autumn, K. & Harmon, L.J. 2017
522 Tempo and mode of performance evolution across multiple independent origins of adhesive
523 toe pads in lizards. *Evolution* **71**, 2344-2358. (doi:10.1111/evo.13318).

524 [28] van der Meij, M.A.A. & Bout, R.G. 2006 Seed husking time and maximal bite force in
525 finches. *Journal Of Experimental Biology* **209**, 3329-3335.

526 [29] Verwaijen, D., Van Damme, R. & Herrel, A. 2002 Relationships between head size, bite
527 force, prey handling efficiency and diet in two sympatric lacertid lizards. *Functional Ecology*
528 **16**, 842-850.

529 [30] Vanhooydonck, B., Herrel, A.Y., Van Damme, R. & Irschick, D.J. 2005 Does Dewlap Size
530 Predict Male Bite Performance in Jamaican Anolis Lizards? *Functional Ecology* **19**, 38-42.

531 [31] Lappin, A., xa, Kristopher, Brandt, Y., Husak, J., xa, F, Macedonia, J., xa, M, et al. 2006
532 Gaping Displays Reveal and Amplify a Mechanically Based Index of Weapon Performance.
533 *The American Naturalist* **168**, 100-113. (doi:10.1086/505161).

534 [32] Hedges, S.B., Marin, J., Suleski, M., Paymer, M. & Kumar, S. 2015 Tree of Life Reveals
535 Clock-Like Speciation and Diversification. *Molecular Biology and Evolution* **32**, 835-845.
536 (doi:10.1093/molbev/msv037).

537 [33] Pagel, M., Meade, A. & Barker, D. 2004 Bayesian estimation of ancestral character
538 states on phylogenies. *Systematic Biology* **53**, 673-684. (doi:10.1080/10635150490522232).

539 [34] van der Meij, M.A.A. & Bout, R.G. 2004 Scaling of jaw muscle size and maximal bite
540 force in finches. *Journal Of Experimental Biology* **207**, 2745-2753.

541 [35] Herrel, A., Podos, J., Huber, S.K. & Hendry, A.P. 2005 Evolution of bite force in Darwin's
542 finches: a key role for head width. *Journal of Evolutionary Biology* **18**, 669-675.

543 [36] Senawi, J., Schmieder, D., Siemers, B. & Kingston, T. 2015 Beyond size – morphological
544 predictors of bite force in a diverse insectivorous bat assemblage from Malaysia. *Functional*
545 *Ecology* **29**, 1411-1420. (doi:10.1111/1365-2435.12447).

546 [37] Brusatte, S.L., Lloyd, G.T., Wang, S.C. & Norell, M.A. 2014 Gradual assembly of avian
547 body plan culminated in rapid rates of evolution across the dinosaur-bird transition. *Current*
548 *Biology* **24**, 2386-2392. (doi:10.1016/j.cub.2014.08.034).

549 [38] Lee, M.S.Y., Cau, A., Naish, D. & Dyke, G.J. 2014 Sustained miniaturization and
550 anatomical innovation in the dinosaurian ancestors of birds. *Science* **345**, 562-566.
551 (doi:10.1126/science.1252243).

552 [39] Puttick, M.N., Thomas, G.H. & Benton, M.J. 2014 High rates of evolution preceded the
553 origin of birds. *Evolution* **68**, 1497-1510. (doi:10.1111/evo.12363).

554 [40] Brusatte, Stephen L., O'Connor, Jingmai K. & Jarvis, Erich D. 2015 The Origin and
555 Diversification of Birds. *Current Biology* **25**, R888-R898.
556 (doi:https://doi.org/10.1016/j.cub.2015.08.003).

557 [41] Brusatte, S.L., Nesbitt, S.J., Irmis, R.B., Butler, R.J., Benton, M.J. & Norell, M.A. 2010 The
558 origin and early radiation of dinosaurs. *Earth-Science Reviews* **101**, 68-100.
559 (doi:10.1016/j.earscirev.2010.04.001).

560 [42] Raftery, A.E. 1996 Hypothesis testing and model selection. In *Markov Chain Monte*
561 *Carlo in Practice* (eds. W.R. Gilks, S. Richardson & D.J. Spiegelhalter), pp. 163-187. London,
562 Great Britain, Chapman & Hall.

563 [43] R Core Development Team. 2011 R: A language and environment for statistical
564 computing. (2.10.1 ed. Vienna, Austria, R Foundation for Statistical Computing.

565 [44] Will, M., Pablos, A. & Stock, J.T. 2017 Long-term patterns of body mass and stature
566 evolution within the hominin lineage. *Royal Society Open Science* **4**.

567 [45] Gignac, P.M. & Erickson, G.M. 2017 The Biomechanics Behind Extreme Osteophagy in
568 *Tyrannosaurus rex*. *Sci Rep-Uk* **7**, 2012. (doi:10.1038/s41598-017-02161-w).

569 [46] Henderson, D.M. & Snively, E. 2004 *Tyrannosaurus en pointe*: allometry minimized
570 rotational inertia of large carnivorous dinosaurs. *Proceedings Of The Royal Society Of*
571 *London Series B-Biological Sciences* **271**, S57-S60.

572 [47] Hutchinson, J.R., Bates, K.T., Molnar, J., Allen, V. & Makovicky, P.J. 2011 A
573 Computational Analysis of Limb and Body Dimensions in *Tyrannosaurus rex* with
574 Implications for Locomotion, Ontogeny, and Growth. *Plos One* **6**, e26037.
575 (doi:10.1371/journal.pone.0026037).

576 [48] Meers, M.B. 2002 Maximum bite force and prey size of *Tyrannosaurus rex* and their
577 relationships to the inference of feeding behaviour. *Historical Biology* **16**, 1-12.

578 [49] Binder, W.J. & Van Valkenburgh, B. 2000 Development of bite strength and feeding
579 behaviour in juvenile spotted hyenas (*Crocuta crocuta*). *Journal of Zoology* **252**, 273-283.

580 [50] Radinsky, L.B. 1981 Evolution of skull shape in carnivores .1. Representative modern
581 carnivores. *Biological Journal of the Linnean Society* **15**, 369-388. (doi:10.1111/j.1095-
582 8312.1981.tb00770.x).

583 [51] Radinsky, L.B. 1981 Evolution of skull shape in carnivores .2. Additional modern
584 carnivores. *Biological Journal of the Linnean Society* **16**, 337-355. (doi:10.1111/j.1095-
585 8312.1981.tb01657.x).

586 [52] Wroe, S., Troy, M., Seebacher, F., Ben, K., Gillespie, A., Mathew, C. & Steve, S. 2003 An
587 alternative method for predicting body mass: The case of the pleistocene marsupial lion.
588 *Paleobiology* **29**, 403-411.

589 [53] Osborn, H.F. 1900 The Geological and Faunal Relations of Europe and America During
590 the Tertiary Period and the Theory of the Successive Invasions of an African Fauna. *Science*
591 **11**, 561-574.

592 [54] Schluter, D. 2000 *The ecology of adaptive radiation*. Oxford, UK, Oxford University
593 Press.

594 [55] Venditti, C., Meade, A. & Pagel, M. 2011 Multiple routes to mammalian diversity.
595 *Nature* **479**, 393-396. (doi:10.1038/nature10516).

596 [56] Grant, B.R. & Grant, P.R. 1993 Evolution of Darwin Finches Caused by a Rare Climatic
597 Event. *Proceedings Of The Royal Society Of London Series B-Biological Sciences* **251**, 111-117.

598 [57] Grant, P.R. & Grant, B.R. 2006 Evolution of character displacement in Darwin's finches.
599 *Science* **313**, 224-226.

600 [58] Lamichhaney, S., Berglund, J., Almen, M.S., Maqbool, K., Grabherr, M., Martinez-Barrio,
601 A., Promerova, M., Rubin, C.-J., Wang, C., Zamani, N., et al. 2015 Evolution of Darwin/'s
602 finches and their beaks revealed by genome sequencing. *Nature* **518**, 371-375.
603 (doi:10.1038/nature14181
604 [http://www.nature.com/nature/journal/v518/n7539/abs/nature14181.html#supplementar](http://www.nature.com/nature/journal/v518/n7539/abs/nature14181.html#supplementary-information)
605 [y-information](http://www.nature.com/nature/journal/v518/n7539/abs/nature14181.html#supplementary-information)).

606 [59] Schluter, D. & Grant, P.R. 1984 Determinants of Morphological Patterns in Communities
607 of Darwin's Finches. *The American Naturalist* **123**, 175-196.

608 [60] Price, T.D., Grant, P.R., Gibbs, H.L. & Boag, P.T. 1984 Recurrent patterns of natural
609 selection in a population of Darwin's finches. *Nature* **309**, 787-789.

610 [61] Pampush, J.D., Scott, J.E., Robinson, C.A. & Delezenne, L.K. 2018 Oblique human
611 symphyseal angle is associated with an evolutionary rate-shift early in the hominin clade.
612 *Journal of Human Evolution*. (doi:<https://doi.org/10.1016/j.jhevol.2018.06.006>).

613 [62] Hublin, J.-J., Neubauer, S. & Gunz, P. 2015 Brain ontogeny and life history in Pleistocene
614 hominins. *Philosophical Transactions of the Royal Society B: Biological Sciences* **370**.

615 [63] Stedman, H.H., Kozyak, B.W., Nelson, A., Thesier, D.M., Su, L.T., Low, D.W., Bridges,
616 C.R., Shrager, J.B., Minugh-Purvis, N. & Mitchell, M.A. 2004 Myosin gene mutation
617 correlates with anatomical changes in the human lineage. *Nature* **428**, 415-418.
618 (doi:http://www.nature.com/nature/journal/v428/n6981/supinfo/nature02358_S1.html).

619 [64] Bruner, E. 2004 Geometric morphometrics and paleoneurology: brain shape evolution
620 in the genus *Homo*. *Journal of Human Evolution* **47**, 279-303.
621 (doi:<http://dx.doi.org/10.1016/j.jhevol.2004.03.009>).

622 [65] McCollum, M.A., Sherwood, C.C., Vinyard, C.J., Lovejoy, C.O. & Schachat, F. 2006 Of
623 muscle-bound crania and human brain evolution: The story behind the MYH16 headlines.
624 *Journal of Human Evolution* **50**, 232-236. (doi:<https://doi.org/10.1016/j.jhevol.2005.10.003>).

625 [66] Perry, G.H., Verrelli, B.C. & Stone, A.C. 2005 Comparative Analyses Reveal a Complex
626 History of Molecular Evolution for Human MYH16. *Molecular Biology and Evolution* **22**, 379-
627 382. (doi:[10.1093/molbev/msi004](https://doi.org/10.1093/molbev/msi004)).

628 [67] Raadsheer, M.C., van Eijden, T.M.G.J., van Ginkel, F.C. & Prah Andersen, B. 1999
629 Contribution of Jaw Muscle Size and Craniofacial Morphology to Human Bite Force
630 Magnitude. *Journal of Dental Research* **78**, 31-42. (doi:[10.1177/00220345990780010301](https://doi.org/10.1177/00220345990780010301)).

631 [68] Orme, D., Freckleton, R., Thomas, G., Petzoldt, T., Fritz, S., Isaac, N. & Pearse, W. 2013
632 caper: Comparative Analyses of Phylogenetics and Evolution in R. R package version 0.5.2.
633 (R package version 0.5.2 ed.

634 [69] Rightmire, G.P. 2004 Brain size and encephalization in early to Mid-Pleistocene *Homo*.
635 *American Journal of Physical Anthropology* **124**, 109-123. (doi:[10.1002/ajpa.10346](https://doi.org/10.1002/ajpa.10346)).

636 [70] DeSilva, J.M. & Lesnik, J.J. 2008 Brain size at birth throughout human evolution: A new
637 method for estimating neonatal brain size in hominins. *Journal of Human Evolution* **55**,
638 1064-1074. (doi:<http://dx.doi.org/10.1016/j.jhevol.2008.07.008>).
639 [71] Isler, K., Christopher Kirk, E., Miller, J.M.A., Albrecht, G.A., Gelvin, B.R. & Martin, R.D.
640 2008 Endocranial volumes of primate species: scaling analyses using a comprehensive and
641 reliable data set. *Journal of Human Evolution* **55**, 967-978.
642 (doi:<http://dx.doi.org/10.1016/j.jhevol.2008.08.004>).
643

FIGURE LEGENDS

Figure 1. Evolution of bite force and its relationship with body mass

Exceptional rates of bite force evolution are shown as a colour gradient (green to gold) on corresponding branches of the phylogenetic tree used in this study, while branches in which no exceptional rates of adaptive changes are detected are in black. Silhouettes highlight clades of interest coloured according to corresponding rates: *Deinonychus antirrhopus*, Maniraptora; *Platyspiza*, branch subtending Passeroidea; *Geospiza fuliginosa*, Thraupidae; Papionin monkey, Anthroidea; and *Homo sapiens*, hominin lineage. Inset, the fitted regression lines from a 5-Group variable-rate regression model accounting for bite point (5-Group+BitePoint VR model) across all MCMC runs are shown in colours corresponding to the five groups of interest: bats, blue; mammals excluding bats, grey; finches, red; dinosaurs excluding finches, turquoise; and reptiles excluding dinosaurs, orange. Significant differences in slopes do not exist between bats and finches as well as between mammals, reptiles and dinosaurs, but significant differences exist between the two sets of groups – i.e., bats/finches and mammals/reptiles/dinosaurs (Table S3). Similarly, slopes in bats and finches are significantly different from the theoretical slope of 0.67 but those in mammals, reptiles and dinosaurs are not (Table S4).

Figure 2. Ancestral reconstruction of bite force across phylogeny and through time

Evolution of bite force with respect to body size while accounting for variable rates show branches and clades with higher amount of change in bite force than expected given ancestral body sizes and phylogenetic positions. Exceptional rates along the branches of the whole tree are shown as a colour gradient (green to gold), with the two clades exhibiting the highest rates (Darwin's finches and hominins) indicated with silhouettes coloured

668 according to corresponding rates (a). Clades in which exceptional rates were detected are
669 highlighted: b, Dinosauria (turquoise) with maniraptorans (skyblue), finches (red) and
670 Darwin's finches (brown); and c, Anthroidea (teal) and hominins (bright green). Insets
671 show subclades of interest (node denoted by a white circle in the whole tree): Darwin's
672 finches amongst the Passeroidea (b) and hominins amongst Hominidae – i.e., great apes (c).
673